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Biofilm formation of *Staphylococcus lugdunensis* strains is associated with clonal lineage and dependent on iron availability

L. Destruel¹, S. Dahyot^{1,2}, X. Argemi^{3,4}, M. Grand¹, G. Prevost³, M. Pestel-Caron^{1,2}

¹Normandie Univ, UNIROUEN, UNICAEN, Inserm UMR 1311 DYNAMICURE, 76000 Rouen, France, ²Normandie Univ, UNIROUEN, UNICAEN, Inserm UMR 1311 DYNAMICURE, CHU Rouen, Department of microbiology, F-76000 Rouen, France, ³VBP EA7290, Fédération de Médecine Translationnelle de Strasbourg, Institut de Bactériologie, Centre Hospitalier Régional Universitaire, Université de Strasbourg, Strasbourg, France, ⁴Maladies Infectieuses et Tropicales, Hôpitaux Universitaires de Strasbourg, Strasbourg, France

Background

Staphylococcus lugdunensis is a virulent coagulase-negative staphylococcal species¹. The ability to form biofilm appears to be one of its main virulence factors², and is promoted by iron-limitation condition³. Nevertheless, little is known about biofilm-associated genotypes.

In this context, the objective of this study was to compare the impact of iron restriction on **fitness** and **biofilm formation** of strains belonging to all clonal complexes (CCs) defined by Multilocus sequence typing⁴ to date.

Materials

Strains

- The study included **49** clinical strains from **different clonal complexes** and **geographical origins** (French and Swedish cities) (Table 1)

Clonal complex	CC1	CC2	CC3	CC4	CC5	CC6	CC7	Singleton*
Number of strains	11	5	11	4	5	5	4	4
Geographical origins	Strasbourg, Rouen, Tours, Kronoberg, Nancy	Strasbourg, Montpellier, Nantes, Nancy, Rouen	Rouen, Strasbourg	Strasbourg, Rouen, Bordeaux	Strasbourg, Bordeaux, Rouen	Kronoberg, Nantes, Strasbourg, Rouen	Strasbourg, Rouen	Rouen, Strasbourg

Table 1: *S. lugdunensis* strains categorized by clonal complexes and geographical origins. *Singleton Sequence Type (ST)

Fitness

- Growth kinetics** over 24 hours performed in triplicates for all strains (Tecan spark®):

- ✓ In **rich media** (Tryptic Soy Broth [TSB])
- ✓ In **iron-restricted condition** (RPMI 1640 with 1% casamino acids)
- ✓ Determination of **generation time** ($G = \ln 2 / \mu$) and **maximal growth rate** (μ) (Figure 1)

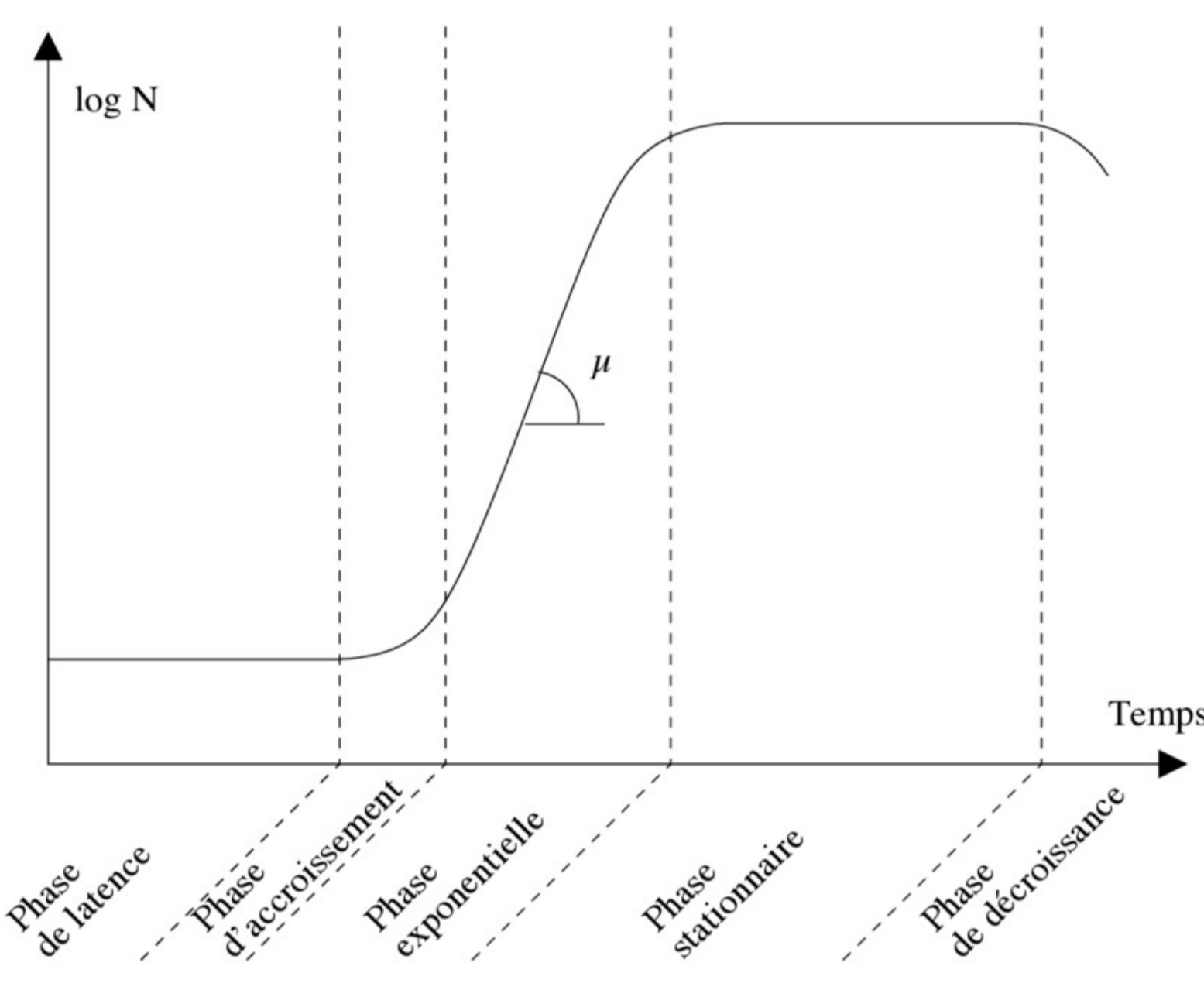


Figure 1 : Diagram of a growth curve

Biofilm formation

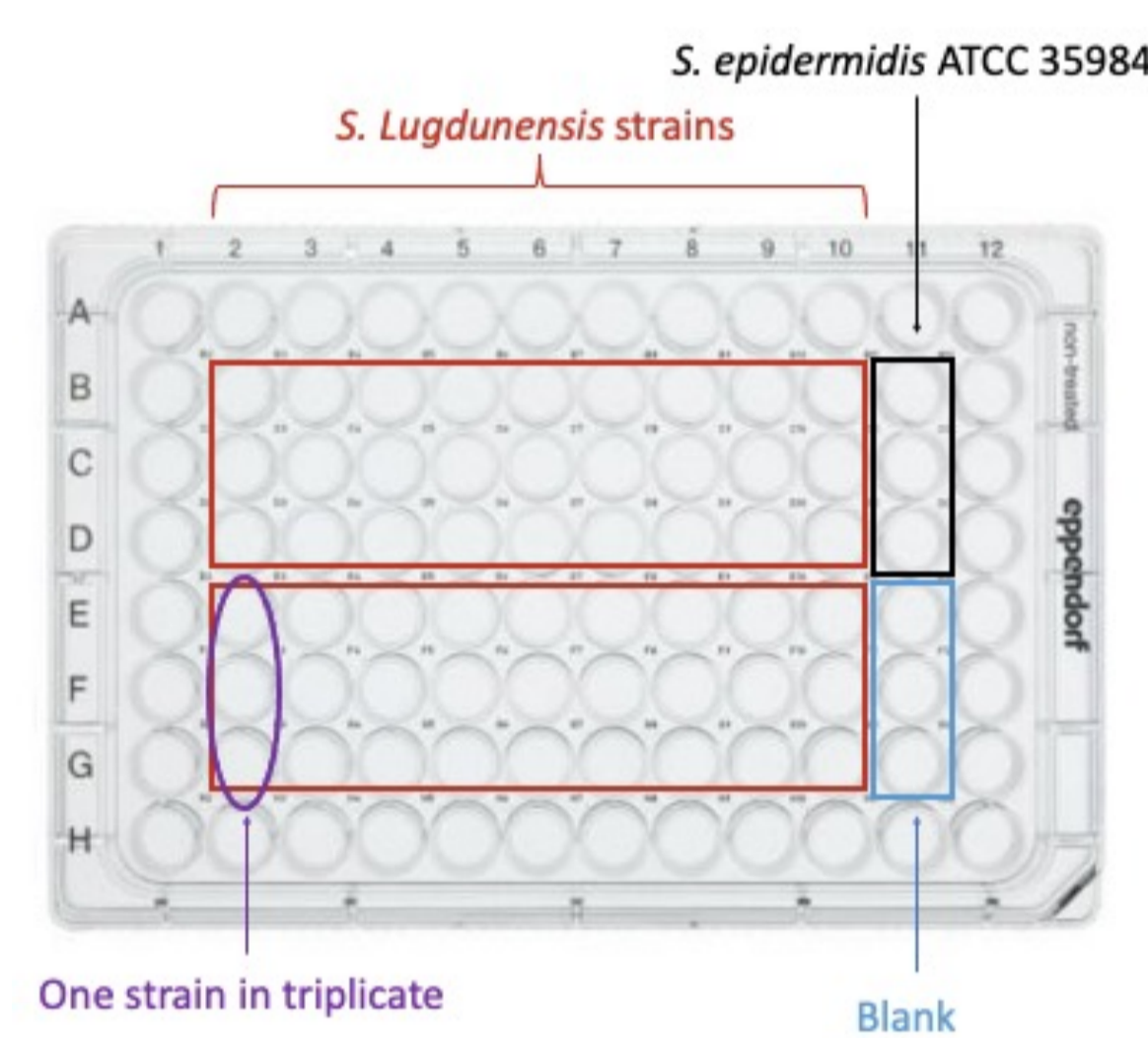


Figure 2: Biofilm formation on polystyrene microtiter plates

- 24h biofilm formation** studied using **crystal violet staining** method on 96-well polystyrene microtiter plates in quadruplicates:

- ✓ In **rich media** (TSB with 1% glucose) for all strains
- ✓ In **iron-restricted condition** for 34 strains (Figure 2)
- ✓ Using *S. epidermidis* ATCC 35984 as biofilm positive control

Results

Fitness

	Generation time (G)	Maximal growth rate (μ)
Rich medium (TSB)	37.34 minutes \pm 4.13	1.139 \pm 0.132
Iron restricted medium (RPMI + 1% CAS)	80.72 minutes \pm 33.14	0.634 \pm 0.20

Table 2: Generation time and maximal growth rate of 49 *S. lugdunensis* strains in rich and iron restricted media

- No difference** in growth either according to the strains or according to the CCs, whatever the medium (Table 2)

Biofilm formation

In rich medium

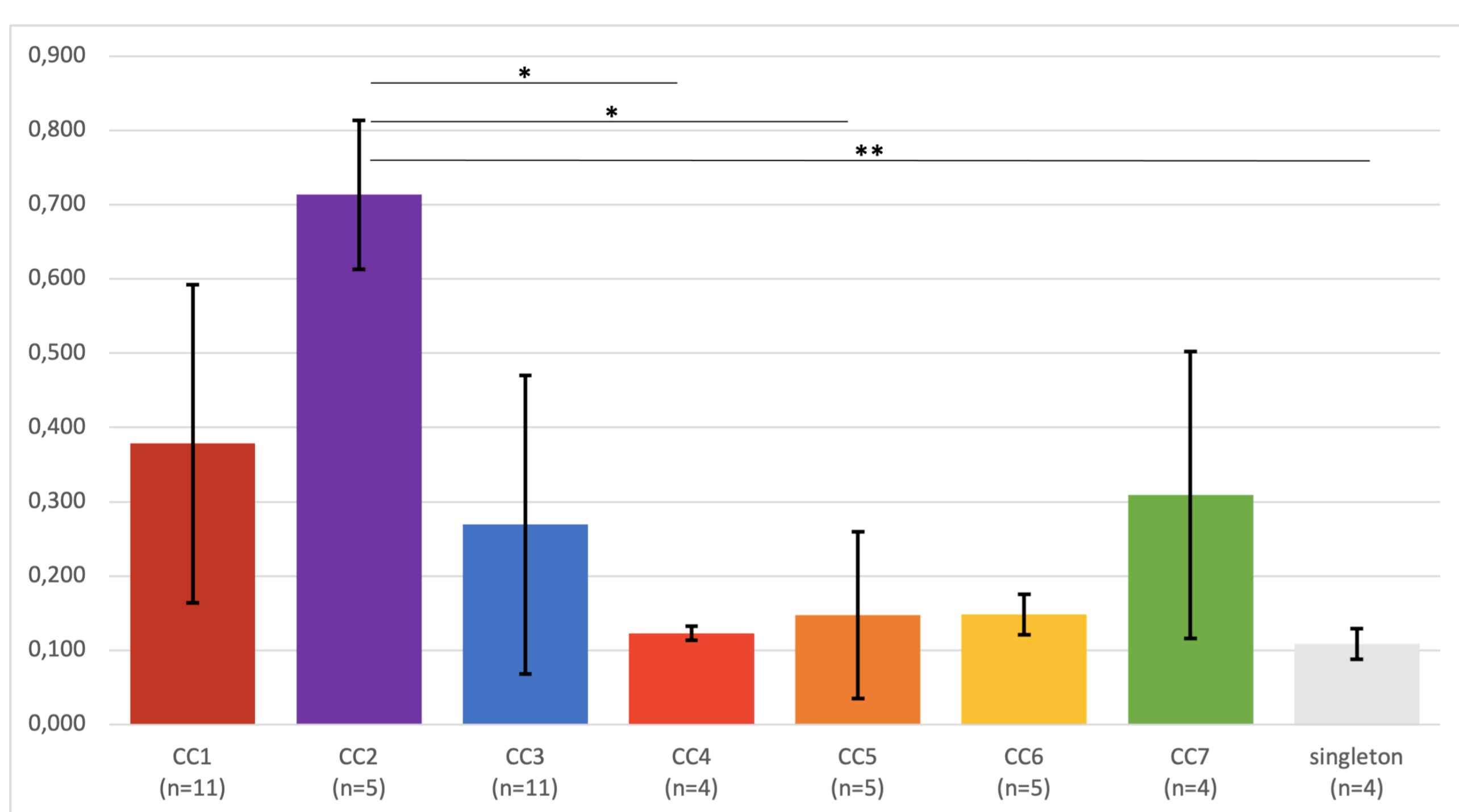


Figure 3: Biofilm formation of 49 *S. lugdunensis* strains after a 24h incubation at 37°C in rich medium (TSB + 1% glucose) according to the CC (OD mean values). Error bars represent the standard deviation within each CC.

- All strains were biofilm producers ($DO_{590nm} > 0,1$) in rich medium after 24h
- CC2** strains produced significantly **more biofilm** than **CC4** ($p = 0.0203$), **CC5** ($p = 0.0215$) and **singleton STs** ($p = 0.0045$) strains (Figure 3)

In iron-restricted conditions

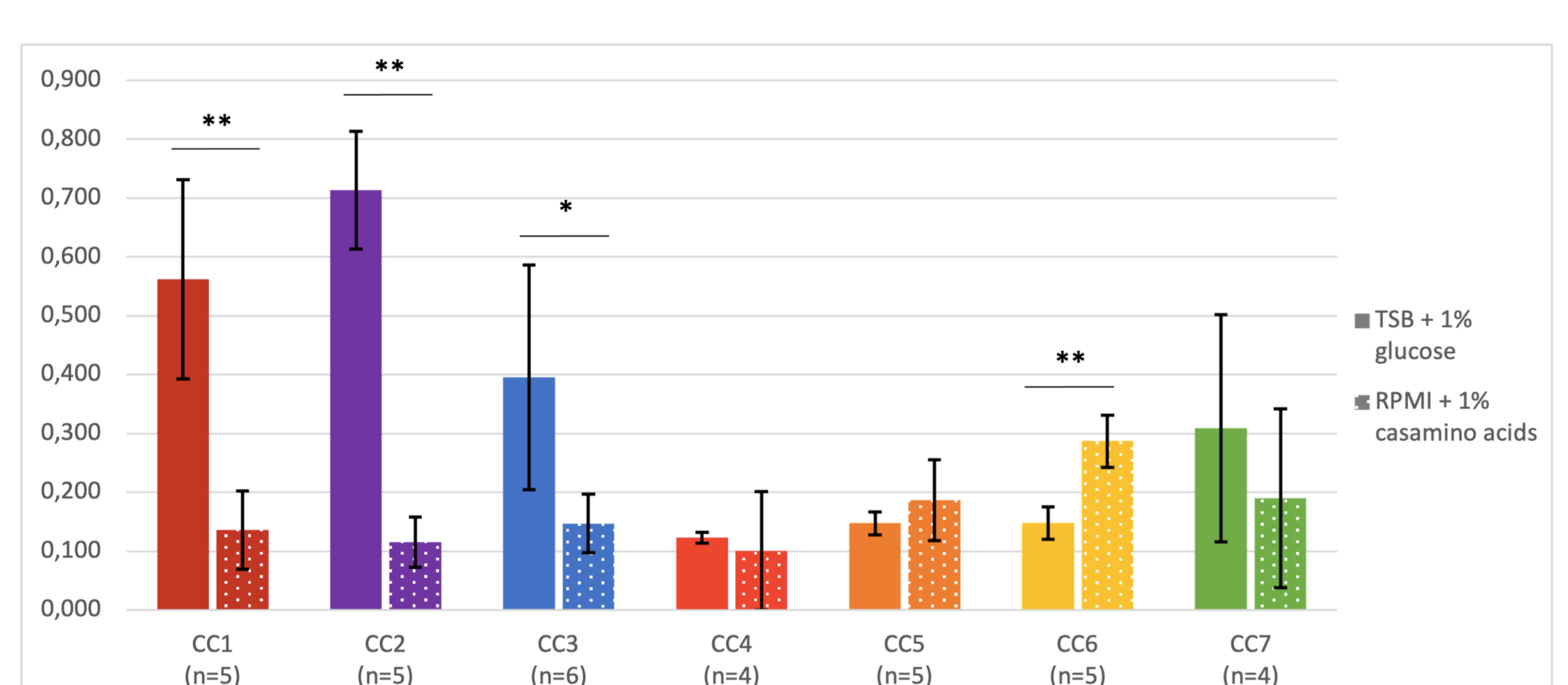


Figure 4: Biofilm formation of 34 *S. lugdunensis* strains after a 24h incubation at 37°C in rich medium (TSB + 1% glucose [full bars]) or iron-restricted medium (RPMI + 1% casamino acids [hatched bars]) according to the CC (OD mean values). Error bars represent the standard deviation within each CC.

- In **iron-restricted conditions**, **CC1** ($p = 0.0079$), **CC2** ($p = 0.0079$) and **CC3** ($p = 0.026$) strains produced significantly **less biofilm** than in rich medium
- On the contrary, **CC6** ($p = 0.0079$) strains were significantly **more productive** (Figure 4)

Conclusions

- This study shows for the first time that the **biofilm-forming ability** of *S. lugdunensis* strains is strongly **correlated with CC affiliation**.
- In addition, two different **CC-dependent behaviors** are observed in an **iron-restricted environment**. These results are to be confirmed with a larger number of strains.

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