

Granins and their derived peptides in normal and tumoral chromaffin tissue: Implications for the diagnosis and prognosis of pheochromocytoma

Marlène Guérin, Johann Guillemot, Erwan Thouennon, Alice Pierre, Fatima-Zohra El-Yamani, Maite Montero-Hadjadje, Christophe Dubessy, Rabia Magoul, Isabelle Lihrmann, Youssef Anouar, et al.

► **To cite this version:**

Marlène Guérin, Johann Guillemot, Erwan Thouennon, Alice Pierre, Fatima-Zohra El-Yamani, et al.. Granins and their derived peptides in normal and tumoral chromaffin tissue: Implications for the diagnosis and prognosis of pheochromocytoma. *Regulatory Peptides*, Elsevier, 2010, 165 (1), pp.21 - 29. 10.1016/j.regpep.2010.06.003 . hal-01706413

HAL Id: hal-01706413

<https://hal-normandie-univ.archives-ouvertes.fr/hal-01706413>

Submitted on 18 Jul 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Granins and their derived peptides in normal and tumoral chromaffin tissue: Implications for the diagnosis and prognosis of pheochromocytoma

Marlène Guérin ^a, Johann Guillemot ^a, Erwan Thouënnon ^a, Alice Pierre ^a, Fatima-Zohra El-Yamani ^b, Maité Montero-Hadjadje ^a, Christophe Dubessy ^a, Rabia Magoul ^b, Isabelle Lihrmann ^a, Youssef Anouar ^a, Laurent Yon ^{a,*}

^a Institut National de la Santé et de la Recherche Médicale (INSERM), U982, Laboratory of Neuronal and Neuroendocrine Differentiation and Communication, European Institute for Peptide Research (IFRMP 23), University of Rouen, Mont-Saint-Aignan, France

^b Laboratory of Neuroendocrinology and Nutritional and Climatic Environment, University Sidi Mohammed Ben Abdellah, Faculty of Sciences Dhar El Mehraz, Fez, Morocco

Contents

1.	Introduction	22
2.	Chromogranin A and derived peptides in pheochromocytoma	23
2.1.	Chromogranin A	23
2.2.	WE-14	24
2.3.	EL35	25
2.4.	Pancreastatin	25
2.5.	Chromacin	25

* Corresponding author. INSERM U982, Laboratory of Neuronal and Neuroendocrine Differentiation and Communication, European Institute for Peptide Research (IFRMP 23), University of Rouen, Faculty of Sciences, Place Emile Blondel, 76821 Mont-Saint-Aignan, France. Tel.: +33 235 14 6945; fax: +33 235 14 6946.

E-mail address: laurent.yon@univ-rouen.fr (L. Yon).

3.	Chromogranin B and derived peptides in pheochromocytoma	25
3.1.	Chromogranin B	25
3.2.	GAWK	25
4.	Secretogranin II and derived peptides in pheochromocytoma	25
4.1.	Secretogranin II	25
4.2.	Secretoneurin	25
4.3.	EM66	26
5.	Other granins and their derived peptides in pheochromocytoma	26
5.1.	SgIII (or 1B1075)	26
5.2.	SgIV (or H1SL-19)	26
5.3.	SgV (or 7B2)	26
5.4.	SgVI (or NESP55)	26
5.5.	SgVII (or VGF) and proSAAS	26
6.	Concluding remarks	26
	References	27

1. Introduction

The granin proteins are widely distributed in secretory granules of endocrine, neuroendocrine and nerve cells [1–3], and have also been described in other secreting cells like exocrine cells, cardiomyocytes or immune system cells [4–6]. At present, the granin family can loosely be divided into two groups, the chromogranins (Cg) and the secretogranins (Sg), distinguishable by the presence of a disulfide-bonded loop at the N-terminus of Cg but not in Sg proteins [7,8]. They constitute a family of acidic glycoproteins whose major members are chromogranin A (CgA), which was first isolated from chromaffin cells of the adrenal medulla [9]; chromogranin B (CgB, previously also called secretogranin I), initially characterized in a rat pheochromocytoma cell line [10]; and secretogranin II (SgII, previously also called chromogranin C), which was originally described in the anterior pituitary [11]. The granin family has further expanded to include other members such as secretogranin III (SgIII or 1B1075) [12], secretogranin IV (SgIV or H1SL-19) [13], secretogranin V (SgV or 7B2) [14], secretogranin VI (SgVI or NESP55) [15], secretogranin VII (SgVII or VGF) [8,16] and proSAAS [17].

Granins are characterized by a high proportion of acidic amino acids (aspartic and glutamic acids) that confers the acidic nature of these proteins. This property enables them to play a role in the formation of secretory vesicles and sequestration of hormones in neuroendocrine cells [18–20]. In addition, granins encompass in their sequence numerous pairs of basic amino acids which are potential cleavage sites for prohormone convertases and which may give rise to bioactive peptides through post-translational proteolytic processing. Indeed, numerous studies have shown that peptides derived from granin maturation occur in various tissues and exhibit autocrine, paracrine, or endocrine activities [21,22]. The ubiquitous distribution of granins in endocrine and neuroendocrine tissues from which they are secreted into the bloodstream, makes granins useful markers of normal and tumoral neuroendocrine cells. In fact, multiple studies have documented the clinical value of detecting granins and their derived peptides in tissues and measuring their circulating levels [7]. In particular, measurement of CgA levels in plasma can be used to diagnose or monitor the progression of neuroendocrine tumors [23]. The highest accuracy has been observed in tumors characterized by an intense secretory activity, although the specificity and sensitivity remain high also in non-functioning tumors [24].

Among neuroendocrine tumors, pheochromocytomas are rare catecholamine-producing neoplasms primarily (80–85%) arising from chromaffin cells of the adrenal medulla or extra-adrenal paraganglia (Fig. 1). Tumors from extra-adrenal location are referred to as paragangliomas. Most of pheochromocytomas occur sporadically but approximately 25% result from germline mutations in 6 genes identified to date. Mutations of the VHL tumor suppressor gene predispose to the development of von Hippel–Lindau (VHL) syndrome, those of NF1

gene to von Recklinghausen's syndrome, and those of the RET proto-oncogene to multiple endocrine neoplasia (MEN) type 2A and 2B [25]. More recently, mutations identified in genes encoding succinate dehydrogenase (SDH) subunits B, C and D have been shown to predispose to familial paraganglioma syndromes [26]. Clinical presentation of pheochromocytoma can vary greatly, with similar signs and symptoms produced by many other clinical conditions. Most but not all the clinical manifestations of pheochromocytoma are due to the indirect actions of secreted catecholamines. Hypertension, tachycardia, pallor, headache and feeling of panic or anxiety, usually dominate the clinical presentation. Metabolic effects include hyperglycemia, lactic acidosis and weight loss. Less common signs and symptoms are nausea, fever and flushing [27]. Despite the increasing availability of molecular diagnostic and prognostic markers, it remains impossible to predict the development of malignant disease, based on histological findings in a resected tumor. Only the presence of metastases of tumoral chromaffin tissue establishes a definite diagnosis of malignant pheochromocytoma [27,28]. It should be noted that the incidence of pheochromocytomas is only 2–8 cases per 1,000,000 subjects and that malignant pheochromocytomas only represent about 10% of all pheochromocytomas.

In this review, we have explored the literature concerning the occurrence of granins in normal and tumoral chromaffin cells, and the

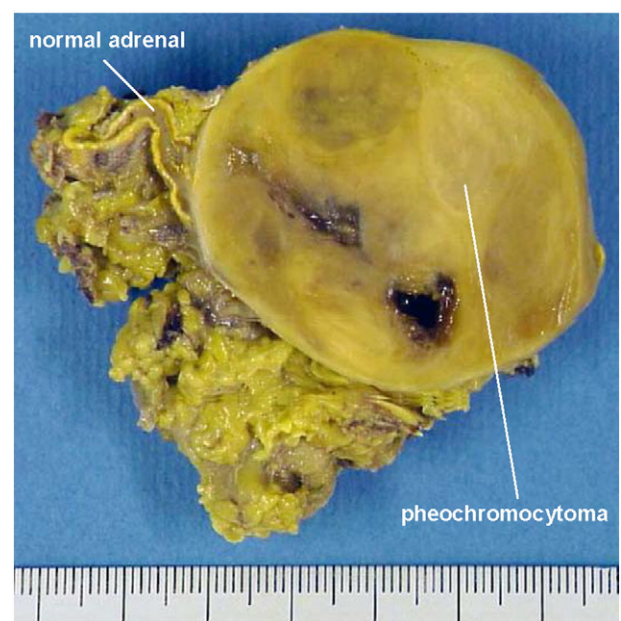


Table 1
Detection of granins and derived peptides in normal and tumoral chromaffin cells, and in circulation. Indication of the test assay sensitivity for the diagnosis of pheochromocytoma.

		Distribution			Test assay sensitivity**
		Normal chromaffin cells		Tumoral chromaffin cells (human pheochromocytoma)	
CgA	CgA	+	h, p, b, m, r	+	71%–100%*
	WE-14	+	h, r	+	84%(n = 25)[66]
	EL35	+	h, r	+	ND
	PST	+	h, pg	+	43%(n = 21)[40]
	chromacin	+	b	+	ND
CgB	CgB	+	h, b, r	+	81%(n = 21)[40]
	GAWK	+	h, b	+	56%GAWK ₁₋₁₇ (n = 9)[77] 33%GAWK ₂₀₋₃₈ (n = 12)
SgII	SgII	+	h, p, b, d, m, r	+	ND
	SN	+	h, b, r	+	78%(n = 9)[87]
	EM66	+	h, b, r	+	92%(n = 12)[66]
SgIII (1B1075)		–		–	ND
SgIV (HISL-19)		+	h	ND	ND
SgV (7B2)		+	h, b, r	+	ND
SgVI (NESP55)		+	h, b, r	+	ND
SgVII (VGF)		+	h, m, r	+	ND
ProSAAS	proSAAS	+	h, m, r	PC12 cells	ND
	SAAS	+	r		ND
	LEN	+	r		ND

A plus sign indicates the presence of the protein, granin-derived peptide and/or messenger RNA; a minus sign indicates absence or undetectable levels. b, bovine; CgA, chromogranin A; CgB, chromogranin B; d, dog; h, human; m, mouse; ND, not determined; p, primate; pg, pig; PST, pancreastatin; r, rat; SgII–VII, secretogranin II–VII; SN, secretoneurin. *The CgA sensitivity test assay for the diagnosis of pheochromocytoma differs according to the following studies: 71% (n = 45) [39]; 83% (n = 29) [36]; 86% (n = 50) [49]; 89% (n = 9) [38]; 90% (n = 21) [40]; 91% (n = 22) [45]; 95% (n = 44) [44]; 100% (n = 15) [41]. **Sensitivity: refers to the probability for a (diagnostic) test to be positive in a population of individuals with the target disorder (tumor, disease...) = true-positive / true-positive + false-negative. Specificity: refers to the probability for a (diagnostic) test to be negative in a population of individuals without the target disorder = true-negative / true-negative + false-positive. False-negative: individuals with the target disorder and negative for the test; False-positive: individuals without the target disorder and positive for the test; True-negative: individuals without the target disorder and negative for the test; True-positive: individuals with the target disorder and positive for the test.

the specificity of the CgA test assay for the diagnosis of pheochromocytoma was only 50% with a positive predictive value of 38%. However, the authors reported that combining CgA with plasma catecholamines provided a lower sensitivity, but excellent specificity, accuracy, and positive predictive value. In clinical practice, there is another major drawback of CgA measurement, *i.e.* elevated levels of CgA are common in patients under treatment with proton pump inhibitors, although no neuroendocrine tumor is present [48,50–52]. It was shown that patients taking proton pump inhibitors alone or in combination with antihypertensive medications had significantly higher CgA levels (mean 635 ng/ml) than subjects without any treatment (mean 174 ng/ml; $P < 0.001$) [48]. Other reasons for non-tumor related increase of CgA are type A gastritis and heart failure [53,54]. Currently, CgA is the only granin whose assay is used routinely by clinicians for the diagnosis of neuroendocrine tumors. However, Stridsberg and collaborators [55] compared the usefulness of three commercial kits (CGA-RIA CT; CIS bio international, Gif-sur-Yvette cedex, France; DAKO Chromogranin A ELISA kit; DAKO A/S, Glostrup, Denmark; and CgA EuroDiagnostica, Malmö, Sweden) in different types of neuroendocrine tumors including pheochromocytomas and showed that, depending on the kit used, the sensitivity of the CgA test varied between 67 and 93%. Thus, in addition to false positive, plasma measurement of CgA may also produce false-negative test results in the diagnosis of neuroendocrine tumors, whose rate may depend on the CgA test used.

Several authors [36,37,41,42,44,56] observed a link between circulating CgA concentration and tumor burden, suggesting that CgA could be a good predictor of tumor size and catecholamine secretion. In contrast, low-secreting or non-functioning chromaffin tumors such as head and neck paragangliomas may go undetected when assessed for CgA. The granin can also be used in the follow-up of patients after resection of tumor and as a prognostic marker. Indeed, it was shown that CgA levels fall to normal shortly after removal of the tumor [24,36,37,42,45,56–58]. Rao and co-workers [59] observed that CgA levels were significantly different in benign vs malignant pheochromocytomas. Indeed, plasma CgA rose progressively from control subjects (48.0 ± 3.0 ng/ml) to patients with benign (188 ± 40.5 ng/ml) or

malignant tumor (2932 ± 960 ng/ml). However, these results were not confirmed in another study which reported similar CgA levels in patients with benign or malignant neoplasms [45]. Nevertheless, Bilek et al. [56] found a significant relationship between plasma CgA concentrations and the PASS score [60] rating the malignancy of pheochromocytoma. Moreover, while performed on a low number of tumors, an immunohistochemical study using six CgA region-specific antibodies reported that the peptide expression pattern differed between benign and malignant pheochromocytomas. The benign neoplastic cells exhibited immunoreactivity to all region-specific antibodies, whereas those of the malignant pheochromocytomas expressed immunoreactivity to only one fragment (CgA_{176–195}) [61]. Finally, CgA may also be indicative of response to therapy in patients with malignant pheochromocytoma. It was shown that, in 5 responders to chemotherapy, there was a significant decline in CgA levels, whereas in 4 nonresponders CgA concentrations did not change, CgA varying longitudinally with tumor response and relapse [59].

2.2. WE-14

WE-14 is a 14 amino-acid long peptide derived from the proteolytic processing of CgA (human CgA_{324–337}) and was first isolated from ileal carcinoid tumor and pheochromocytoma tissues [62,63] (Fig. 2). WE-14 is widely distributed in neuroendocrine tissues including rat and human chromaffin cells [64]. Moreover, WE-14 immunostaining was also observed in tumoral chromaffin cells [65]. HPLC analysis combined with RIA detection demonstrated that WE-14 was present as a single immunoreactive peak in pheochromocytoma extracts. Measurement of circulating levels of WE-14 revealed significantly higher concentrations in patients with pheochromocytoma compared to healthy volunteers, the median preoperative level of WE-14 being about 4-fold higher [66]. *In vitro*, using cultured cells from a benign pheochromocytoma, it was demonstrated that metoclopramide, a gastroprokinetic and antiemetic agent, induced a concentration-dependent increase in WE-14 production with an EC₅₀ reaching 3.84×10^{-8} M, and E_{max} of $78 \pm 12\%$ [67]. Moreover, the WE-14 response to metoclopramide was inhibited by

GR 113808 (10^{-7} M), a 5-HT₄ receptor antagonist. Cisapride, a 5-HT₄ receptor agonist, and dopamine were also shown to exert stimulatory effects on WE-14 release from pheochromocytoma cells [67]. These observations indicate that the levels of CgA-derived peptides may vary in pheochromocytomas in response to therapeutic interventions.

2.3. EL35

EL35 (human CgA₄₀₂₋₄₃₆) is a well-conserved peptide derived from the C-terminal domain of CgA (Fig. 2). Consistent with WE-14, EL35-like labeling is found in chromaffin cells of the rat and human normal adrenal glands as well as in benign pheochromocytomas [65]. However, HPLC characterization revealed only minute amounts of authentic EL35 in pheochromocytoma extracts while other EL35-immunoreactive forms were observed. The identity of this immunoreactive material was not determined but it should be mentioned that EL35 possesses an internal pair of basic residues which could be cleaved to give rise to shorter peptides. Therefore, the proteolytic processing of CgA to generate this peptide could be altered during tumorigenesis [65].

2.4. Pancreastatin

Pancreastatin (PST, human CgA₂₅₀₋₃₀₁) is a 52 amino-acid peptide located in the internal part of the CgA sequence, showing a low sequence similarity in vertebrate species [21] (Fig. 2). Using an antibody directed against synthetic porcine PST, Bishop et al. [68] reported the occurrence of PST-like immunoreactivity in a series of human neuroendocrine tumors. PST levels have been determined through RIA measurement in pheochromocytomas (from 8.0 to 69.0 pmol/g). However, no labeling could be observed by immunocytochemistry in the same study. This discrepancy was ascribed to the fact that the antiserum to PST was raised against the porcine peptide. The development of a specific assay for the measurement of circulating human PST allowed Stridsberg and Husebye [40] to investigate the usefulness of this peptide for the diagnosis of patients with pheochromocytoma. They found a low diagnostic sensitivity for this tumor (43%), since plasma PST levels were elevated in only nine out of twenty one patients (Table 1). This finding does not speak for the usefulness of PST as a marker for the diagnosis of pheochromocytoma.

2.5. Chromacin

Using antibodies directed against six regions of the CgA molecule including the chromacin peptide (human CgA₁₇₆₋₁₉₇) (Fig. 2), Portela-Gomes et al. [61] described a positive labeling for this CgA region in different neuroendocrine tumors studied including pheochromocytomas. Moreover, semi-quantitative analysis revealed a more intense chromacin-specific labeling in benign than in malignant pheochromocytomas, but this finding was not confirmed by other biochemical analyses.

3. Chromogranin B and derived peptides in pheochromocytoma

3.1. Chromogranin B

CgB is an abundant secretory protein found in a large variety of endocrine cells and neurons [69] (Fig. 2). CgB has been described in chromaffin cells of different mammalian species [70,71] and in pheochromocytoma extracts [72]. In benign pheochromocytomas, CgB was consistently detected, whereas in two out of three malignant tumors, the granin protein was absent [73]. In pheochromocytomas, while variations in the expression pattern occurred, no significant semi-quantitative differences were noted between benign and malignant tumors [74]. Nevertheless, in all malignant pheochromocytomas, the antibodies raised against the C-terminal region of CgB revealed a noticeable population of immunoreactive large spindle-shaped tumor

cells, characterized by elongated processes. Interestingly, this cell type was only observed in one out of 25 benign tumors. Therefore, the authors suggested that the use of antibodies raised against epitopes of the C-terminal region of CgB could facilitate the diagnosis of malignant pheochromocytomas [74]. Stridsberg and Husebye [40] developed a specific assay for the measurement of circulating CgB, in order to investigate the usefulness of this protein for the diagnosis of patients with pheochromocytoma. They found a diagnostic sensitivity of the CgB test assay of 81%, the granin level being elevated in 17 out of 21 patients with pheochromocytoma (Table 1). While the CgA levels fell to normal shortly after the tumor was removed, the CgB levels decreased towards normal over the course of several days [40].

3.2. GAWK

GAWK (human CgB₄₂₀₋₄₉₃) is a 74 amino-acid peptide originally isolated from human pituitary gland extracts [75,76] (Fig. 2). Bishop et al. [77] have shown that GAWK-like immunoreactivity is present in chromaffin cells of the human adrenal medulla. Using two different antibodies (directed against the GAWK₁₋₁₇ and GAWK₂₀₋₃₈ fragments), a consistent positive immunoreactive staining was seen in all pheochromocytomas studied (n = 6) [78]. In contrast, only antibodies against GAWK₂₀₋₃₈ yielded an immunostaining in cells of normal adrenal medulla. RIA quantification revealed that GAWK₂₀₋₃₈ and GAWK₁₋₁₇ concentrations were about 15-fold higher in tumoral compared to normal chromaffin cells [78]. The GAWK levels were at least 10 times higher in pheochromocytomas than in any other GAWK-producing tumor. While circulating GAWK₁₋₁₇ levels were systematically more elevated in pheochromocytoma patients than in healthy volunteers (121 ± 66 pmol/l vs 7.0 ± 1.1 pmol/l), this was not the case for the GAWK₂₀₋₃₈ peptide (114 ± 19 pmol/l vs 74 ± 5.0 pmol/l) [78] (Table 1).

4. Secretogranin II and derived peptides in pheochromocytoma

4.1. Secretogranin II

SgII was observed in human adrenomedullary tissue and pheochromocytomas [71,73,79]. In the tumors, while variations in the staining pattern with different SgII antibodies were observed, no significant semi-quantitative differences were noted between benign and malignant neoplasms [74]. However, these authors described SgII-immunoreactive large spindle-shaped pheochromocytoma cells preferentially labeled in malignant tumors, as previously observed with CgB labeling. Guillemot et al. [80] showed that SgII mRNA was overexpressed in pheochromocytoma compared to non-tumoral chromaffin tissue and in benign compared to malignant tumors. In addition, western blot quantification revealed that the SgII protein was significantly more abundant in benign than in malignant neoplasms. Using region-specific antisera, Stridsberg et al. [81] showed differential occurrence of SgII-derived fragments in the plasma of pheochromocytoma patients.

4.2. Secretoneurin

The SgII-derived peptide SN (human SgII₁₅₂₋₁₈₄) is a highly conserved 33 amino-acid peptide [82,83] (Fig. 2) that was initially described in the rat and human adrenal medulla [84,85]. It was shown that SgII processing to yield SN was limited in the adrenal medulla and pituitary compared to other organs. Ischia et al. [86] determined the levels of SN in the serum of healthy subjects and patients suffering of pheochromocytoma. They first showed that the mean serum concentration of SN in 42 controls was 22.2 ± 1.1 fmol/ml (range 9.1–37.9). They observed that SN levels were age-dependent with higher concentrations in newborns followed by a continuous decline until 12 years, where adult levels were already reached. It was shown

that the contribution of the adrenal gland to serum SN-immunoreactivity was limited since, in patients with unilateral or bilateral adrenalectomy, no significant reduction in serum SN was observed [86]. In patients with pheochromocytoma, serum SN levels were elevated up to 5-fold (99.9 ± 25.6 fmol/ml) with only few patients within the normal range (Table 1). After surgical resection of the tumor, SN values returned to normal (18.3 ± 2.9 fmol/ml) [86]. Similar results were obtained by Stridsberg et al. [81], although these authors found concentrations of SN 10-fold higher in healthy volunteers with a reference range of 220 fmol/ml and concentrations up to 5400 fmol/ml in patients with pheochromocytoma. These differences could probably be ascribed to the different biological tools and methods used. However, these studies agree on the significant elevation of SN in pheochromocytoma patients.

4.3. EM66

EM66 (human SgII₁₈₇₋₂₅₂) is a 66 amino-acid long peptide that was initially identified by us (Fig. 2) [87,88]. We demonstrated that EM66 was produced in chromaffin cells of the adult and fetal human adrenal gland [89]. Subsequently, the presence of EM66-immunoreactivity was shown in rat and bovine adrenomedullary cells [89,90] (Table 1). Immunohistochemical analysis demonstrated the occurrence of authentic EM66 in both benign and malignant pheochromocytomas [91]. RIA quantification revealed that a low EM66 concentration in pheochromocytoma was associated with a malignant status of this tumor [80,91]. It was also shown that pheochromocytoma cells in primary culture had the ability to release high basal levels of EM66, suggesting that, *in vivo*, the peptide is secreted from the tumor in the general circulation [92]. In accordance, RIA determination revealed that plasma levels of EM66 were significantly higher in patients with pheochromocytoma compared to healthy volunteers (median 2.6, [1.9–3.7] ng/ml vs 26.9 [7.3–44] ng/ml, respectively) and returned to normal values after removal of the tumor [92]. In contrast, urinary EM66 levels in patients with pheochromocytoma were not significantly different from those of controls. Stridsberg et al. [81] found no change of EM66-like concentrations in patients with pheochromocytoma (see Section 4.1). This discrepancy may be ascribed to the antiserum used recognizing SgII₂₂₅₋₂₄₂ *i.e.* C-terminal part of EM66, and/or to the low number of cases studied ($n = 3$). As observed for the WE-14 peptide (see Section 2.2), metoclopramide, cisapride and dopamine dose-dependently stimulated EM66 release from cultured pheochromocytoma cells [67], again indicating that the levels of granin peptides are prone to variation under therapeutic treatment, at least with drugs targeting aminergic systems.

5. Other granins and their derived peptides in pheochromocytoma

While less studied, secretogranins III–VII and proSAAS also belong to the granin family and are characterized by numerous pairs of basic amino acids which may serve as potential cleavage sites for processing by the co-stored prohormone converting enzymes PC1/3 and PC2 in secretory granules [8] (Fig. 2). However, their presence and their use as circulating markers for pheochromocytomas have not been extensively explored (Table 1).

5.1. SgIII (or 1B1075)

SgIII is distinct from other granins as it is not found either in the adrenal gland or in pheochromocytomas [81].

5.2. SgIV (or H1SL-19)

SgIV is a protein expressed by neuronal and peptide hormone-producing cells which shares many biochemical and molecular key features with the granin family [13]. Few studies reported the presence of SgIV in

human adrenal medulla and in pheochromocytoma [13,93,94]. Immunoelectron microscopic analysis revealed SgIV immunoreactivity in secretory granules, which was apparently more pronounced in malignant compared with benign pheochromocytomas [94]. No data are available concerning circulating SgIV in pheochromocytoma patients (Table 1).

5.3. SgV (or 7B2)

7B2 is an acidic protein contained in most neuroendocrine cells. The presence of 7B2 was demonstrated in chromaffin tissue of the normal adrenal gland [95–98] and also in pheochromocytoma [98,99] (Table 1). In human, Natori and co-workers [100] determined plasma 7B2 levels in healthy volunteers and in patients with pheochromocytoma. They measured significantly higher plasma 7B2 concentrations in patients with pheochromocytoma than those in age-matched normal subjects (221 ± 82.8 ng/l vs 55.8 ± 1.2 ng/l).

5.4. SgVI (or NESP55)

NESP55 (neuroendocrine secretory protein of MW 55,000) is a soluble, acidic, heat-stable secretory protein [101] (Fig. 2) which was described in the rat and bovine adrenal medulla with a preferential localization in adrenaline-synthesizing cells [15,102]. NESP55 immunoreactivity was also shown in pheochromocytomas, with the same strong labeling pattern in both benign and malignant tumors [103–105] (Table 1).

5.5. SgVII (or VGF) and proSAAS

SgVII/VGF and proSAAS represent the most recently characterized members of the granin family (Fig. 2). VGF was originally identified as a neurotrophin-regulated gene product in PC12 cells [16]. ProSAAS was identified as the precursor of several peptides that accumulate in the obese, carboxypeptidase E-deficient *Cpefat/fat* mouse, and was shown to inhibit prohormone convertase 1 activity [17]. VGF and proSAAS are stored in and released from secretory vesicles and are expressed widely in nervous, endocrine, and neuroendocrine tissues [17,106,107]. Variable proVGF-immunoreactive fragments were observed in human adult adrenal medulla and in pheochromocytomas [108]. ProSAAS mRNA is primarily expressed in the brain but also occurs in endocrine organs and cells including the adrenal gland and the rat pheochromocytoma PC12 cell line [17] (Table 1). Two proSAAS-derived peptides (SAAS and LEN) were detected in the rat adrenal medulla [109] (Fig. 2).

6. Concluding remarks

Malignant pheochromocytomas are orphan diseases for which there exist a difficulty for the management and a disparity in clinical practices. To date, the universally admitted criterion of malignancy for a pheochromocytoma is the occurrence of metastasis [110,111]. Therefore, numerous efforts have been undertaken to discover reliable tools for the early diagnosis and prognosis of malignant pheochromocytomas. While pathological analysis utilizing multiple histological criteria appears to hold some value for distinguishing benign from malignant disease, these evaluations are not practiced routinely as predictors of malignancy [60,112,113]. Among pheochromocytoma susceptibility genes, it is now established that SDHB mutation is associated with the highest risk of malignancy [114]. It has also been shown that malignant pheochromocytomas are characterized by an enhanced production of noradrenaline and that extra-adrenal (paragangliomas) tumors secrete dopamine [115]. A global and promising approach of gene expression profiling of benign and malignant pheochromocytomas, has allowed to identify a set of genes that could discriminate the two types of tumors [114,116]. As indicated in the present review, except SgIII, all members

of the granin family and several of their derived peptides have been shown to be present in normal chromaffin cell and in pheochromocytomas (Table 1). However, to date, CgA is the only granin currently used for the diagnosis of chromaffin cell tumors; a positive CgA immunostaining indicating the neuroendocrine nature of the neoplasm in general, without specificity for pheochromocytoma. Currently, besides CgA, other biochemical tests used for the diagnosis of pheochromocytoma are the measurement of plasma and urinary catecholamine metabolites (metanephrine and normetanephrine). While they offer a high sensitivity, these tests also have limitations. To date, a biochemical test alone is obviously not sufficient for the screening of pheochromocytoma. False-negative and false-positive results using biological tests remain a problem, leading to cost-effective and time-consuming additional imaging examinations [117,118]. It is therefore tempting to propose that the measurement of CgA and metanephrine levels should be combined with the determination of other granins or granin-derived peptides for the diagnosis/prognosis of pheochromocytoma.

CgA levels are correlated with tumor mass since malignant pheochromocytomas which are often more bulky than benign ones display higher CgA concentrations [43]. Thus, CgA may serve as a prognostic marker in certain conditions but there are several restrictions such as the existence of malignant non-secreting chromaffin tumors of the head and neck. Other drawbacks of CgA limit the clinical use of the protein (false-negative or false-positive test results) and therefore, there is a need to identify other granin or granin-derived peptides for the diagnosis and prognosis of pheochromocytomas. In this respect, CgB did not seem to have a better diagnostic sensitivity than CgA but was shown to be a complementary tool for the management of neuroendocrine tumors, since its concentrations were not elevated in patients with renal failure or under treatment with proton pump inhibitors [57]. It was also postulated by the group of Stridsberg and Portela-Gomes that the use of antibodies raised against epitopes of the C-terminal region of CgB could facilitate the diagnosis of malignant pheochromocytoma [74]. Our data showed that SgII mRNAs were down-regulated in malignant compared to benign pheochromocytomas and that malignancy was associated with reduced levels of SgII and SgII-derived peptides, in accordance with the low concentrations of EM66 found in malignant tumors [80,91]. Taken together, these observations strengthen the notion that granins other than CgA and their derived peptides may provide new diagnostic and prognostic markers of pheochromocytoma.

An important issue raised by A. Tischler, is to define more precisely the term "malignant" concerning pheochromocytoma/paraganglioma since the potential lethality of extensive local invasion is ignored by a definition of malignancy based only on metastasis [119]. He suggested that pheochromocytoma and extra-adrenal paraganglioma could be classified as "non-invasive", "minimally invasive" and "extensively invasive", in order to stratify the risk of metastasis and aggressiveness for a better management of patient treatment [119]. In this regard, the development of additional markers derived from granins and their peptides used solely or in combination, *i.e.* through array of different peptide-recognizing antibodies, may provide a mean to accurately discriminate different pheochromocytoma subtypes. However, a precise description of the relative affinity of each antibody raised against a granin-derived peptide towards the peptide itself and its precursor will be needed because this important information is not always available. Other challenges to the diagnosis and prognosis of pheochromocytoma will be to detect tumors that do not contain or produce excess catecholamines, metanephrines or CgA. Concerning the prognostic value of histopathology of pheochromocytoma tumors, to date there is no consensus on the reliability of the PASS score [60]. Further studies should combine histological (PASS), immunohistochemical (Ki67) and biochemical (adrenergic/noradrenergic, granins) features to predict metastatic potential of tumors [113,120,121]. However, it should be kept in mind that these different markers would carry different weights depending on the adrenal or extra-adrenal localization and the genetic background of the tumor [119]. In addition, chromaffin tumors are rare, heterogeneous, and further work aimed at

identifying new predictive factors of malignancy should be performed with large and well characterized cohorts of patients. Finally, *in vitro* experiments, performed on tumor cell lines, suggest that CgA may affect tissue remodeling and tumor growth [122]. Therefore, in addition to their potential use as predictors for the diagnosis and prognosis, granin-derived peptides could also be considered for use in therapeutic strategies for the treatment of pheochromocytoma and other neuroendocrine tumors.

References

- Rosa P, Gerdes HH. The granin protein family: markers for neuroendocrine cells and tools for the diagnosis of neuroendocrine tumors. *J Endocrinol Invest* 1994;17:207-25.
- Fischer-Colbrie R, Laslop A, Kirchmair R. Secretogranin II: molecular properties, regulation of biosynthesis and processing to the neuropeptide secretoneurin. *Prog Neurobiol* 1995;46:49-70.
- Winkler H, Fischer-Colbrie R. The chromogranins A and B: the first 25 years and future perspectives. *Neuroscience* 1992;49:497-528.
- Steiner HJ, Weiler R, Ludescher C, Schmid KW, Winkler H. Chromogranins A and B are co-localized with atrial natriuretic peptides in secretory granules of rat heart. *J Histochem Cytochem* 1990;38:845-50.
- Tasiemski A, Hammad H, Vandenbulcke F, Breton C, Bilfinger TJ, Pestel J, Salzet M. Presence of chromogranin-derived antimicrobial peptides in plasma during coronary artery bypass surgery and evidence of an immune origin of these peptides. *Blood* 2002;100:553-9.
- Saruta J, Tsukinoki K, Sasaguri K, Ishii H, Yasuda M, Osamura YR, Watanabe Y, Sato S. Expression and localization of chromogranin A gene and protein in human submandibular gland. *Cells Tissues Organ* 2005;180:237-44.
- Taupenot L, Harper KL, O'Connor DT. The chromogranin-secretogranin family. *N Engl J Med* 2003;348:1134-49.
- Helle KB. The granin family of uniquely acidic proteins of the diffuse neuroendocrine system: comparative and functional aspects. *Biol Rev Camb Philos Soc* 2004;79:769-94.
- Banks P, Helle K. The release of protein from the stimulated adrenal medulla. *Biochem J* 1965;97:40C-1C.
- Lee RW, Huttner WB. Tyrosine-O-sulfated proteins of PC12 pheochromocytoma cells and their sulfation by a tyrosylprotein sulfotransferase. *J Biol Chem* 1983;258:11326-34.
- Rosa P, Hille A, Lee RW, Zanini A, De Camilli P, Huttner WB. Secretogranins I and II: two tyrosine-sulfated secretory proteins common to a variety of cells secreting peptides by the regulated pathway. *J Cell Biol* 1985;101:1999-2011.
- Ottiger HP, Battenberg EF, Tsou AP, Bloom FE, Sutcliffe JG. 1B1075: a brain- and pituitary-specific mRNA that encodes a novel chromogranin/secretogranin-like component of intracellular vesicles. *J Neurosci* 1990;10:3135-47.
- Krisch K, Buxbaum P, Horvat G, Krisch I, Neuhold N, Ulrich W, Srikanta S. Monoclonal antibody HSL-19 as an immunocytochemical probe for neuroendocrine differentiation. Its application in diagnostic pathology. *Am J Pathol* 1986;123:100-8.
- Iguchi H, Chan JS, Seidah NG, Chretien M. Tissue distribution and molecular forms of a novel pituitary protein in the rat. *Neuroendocrinology* 1984;39:453-8.
- Ischia R, Lovisetti-Scamihorn P, Hogue-Angeletti R, Wolkersdorfer M, Winkler H, Fischer-Colbrie R. Molecular cloning and characterization of NESP55, a novel chromogranin-like precursor of a peptide with 5-HT1B receptor antagonist activity. *J Biol Chem* 1997;272:11657-62.
- Levi A, Eldridge JD, Paterson BM. Molecular cloning of a gene sequence regulated by nerve growth factor. *Science* 1985;229:393-5.
- Fricker LD, McKinzie AA, Sun J, Curran E, Qian Y, Yan L, Patterson SD, Courchesne PL, Richards B, Levin N, Mzhavria N, Devi LA, Douglass J. Identification and characterization of proSAAS, a granin-like neuroendocrine peptide precursor that inhibits prohormone processing. *J Neurosci* 2000;20:639-48.
- Chanat E, Huttner WB. Milieu-induced, selective aggregation of regulated secretory proteins in the trans-Golgi network. *J Cell Biol* 1991;115:1505-19.
- Iacangelo AL, Eiden LE. Chromogranin A: current status as a precursor for bioactive peptides and a granulogenic/sorting factor in the regulated secretory pathway. *Regul Pept* 1995;58:65-88.
- Kim T, Tao-Cheng JH, Eiden LE, Loh YP. Chromogranin A, an "on/off" switch controlling dense-core secretory granule biogenesis. *Cell* 2001;106:499-509.
- Montero-Hadjadj M, Vaingankar S, Elias S, Tostivint H, Mahata SK, Anouar Y. Chromogranins A and B and secretogranin II: evolutionary and functional aspects. *Acta Physiol (Oxf)* 2008;192:309-24.
- Zhao E, Zhang D, Basak A, Trudeau VL. New insights into granin-derived peptides: evolution and endocrine roles. *Gen Comp Endocrinol* 2009;164:161-74.
- Ferrari L, Seregni E, Bajetta E, Martinetti A, Bombardieri E. The biological characteristics of chromogranin A and its role as a circulating marker in neuroendocrine tumours. *Anticancer Res* 1999;19:3415-27.
- Kimura N, Miura W, Noshiro T, Mizunashi K, Hanew K, Shimizu K, Watanabe T, Shibukawa S, Sohn HE, Abe K, Miura Y, Nagura H. Plasma chromogranin A in pheochromocytoma, primary hyperparathyroidism and pituitary adenoma in comparison with catecholamine, parathyroid hormone and pituitary hormones. *Endocr J* 1997;44:319-27.
- Thouñnon E, Elkahloun AG, Guillemot J, Gimenez-Roqueplo AP, Bertherat J, Pierre A, Ghzili H, Grumolato L, Muresan M, Klein M, Lefebvre H, Vaudry

- H, Plouin PF, Yon L, Anouar Y. Identification of potential gene markers and insights into the pathophysiology of pheochromocytoma malignancy. *J Clin Endocrinol Metab* 2007;92:4865–72.
- [26] Burnichon N, Rohmer V, Amar L, Herman P, Lebouilleux S, Darrouzet V, Niccoli P, Gaillard D, Chabrier G, Chabolle F, Coupier I, Thieblot P, Lecomte P, Bertherat J, Wion-Barbot N, Murat A, Venisse A, Plouin PF, Jeunemaitre X, Gimenez-Roqueplo AP. The succinate dehydrogenase genetic testing in a large prospective series of patients with paragangliomas. *J Clin Endocrinol Metab* 2009;94:2817–27.
- [27] Lenders JW, Eisenhofer G, Mannelli M, Pacak K. Pheochromocytoma. *Lancet* 2005;366:665–75.
- [28] Scholz T, Schulz C, Klose S, Lehnert H. Diagnostic management of benign and malignant pheochromocytoma. *Exp Clin Endocrinol Diab* 2007;115:155–9.
- [29] O'Connor DT. Chromogranin: widespread immunoreactivity in polypeptide hormone producing tissues and in serum. *Regul Pept* 1983;6:263–80.
- [30] O'Connor DT, Burton D, Defetos LJ. Chromogranin A: immunohistology reveals its universal occurrence in normal polypeptide hormone producing endocrine glands. *Life Sci* 1983;33:1657–63.
- [31] O'Connor DT, Burton D, Defetos LJ. Immunoreactive human chromogranin A in diverse polypeptide hormone producing human tumors and normal endocrine tissues. *J Clin Endocrinol Metab* 1983;57:1084–6.
- [32] O'Connor DT, Bernstein KN. Radioimmunoassay of chromogranin A in plasma as a measure of exocytotic sympathoadrenal activity in normal subjects and patients with pheochromocytoma. *N Engl J Med* 1984;311:764–70.
- [33] Dillen L, De Block J, Van Lear L, De Potter W. Enzyme-linked immunosorbent assay for chromogranin A. *Clin Chem* 1989;35:1934–8.
- [34] Degorce F, Goumon Y, Jacquemart L, Vidaud C, Bellanger L, Pons-Anicet D, Seguin P, Metz-Boutigue MH, Aunis D. A new human chromogranin A (CgA) immunoradiometric assay involving monoclonal antibodies raised against the unprocessed central domain (145–245). *Br J Cancer* 1999;79:65–71.
- [35] Cleary S, Phillips JK, Huynh TT, Pacak K, Fliedner S, Elkahoul AG, Munson P, Worrell RA, Eisenhofer G. Chromogranin A expression in pheochromocytomas associated with von Hippel–Lindau syndrome and multiple endocrine neoplasia type 2. *Horm Metab Res* 2007;39:876–83.
- [36] Hsiao RJ, Parmer RJ, Takiyyuddin MA, O'Connor DT. Chromogranin A storage and secretion: sensitivity and specificity for the diagnosis of pheochromocytoma. *Medicine (Baltimore)* 1991;70:33–45.
- [37] Hsiao RJ, Neumann HP, Parmer RJ, Barbosa JA, O'Connor DT. Chromogranin A in familial pheochromocytoma: diagnostic screening value, prediction of tumor mass, and post-resection kinetics indicating two-compartment distribution. *Am J Med* 1990;88:607–13.
- [38] Nobels FR, Kwekkeboom DJ, Coopmans W, Schoenmakers CH, Lindemans J, De Herder WW, Krenning EP, Bouillon R, Lamberts SW. Chromogranin A as serum marker for neuroendocrine neoplasia: comparison with neuron-specific enolase and the alpha-subunit of glycoprotein hormones. *J Clin Endocrinol Metab* 1997;82:2622–8.
- [39] Boomsma F, Bhaggoo UM, Man in 't Veld AJ, Schalekamp MA. Sensitivity and specificity of a new ELISA method for determination of chromogranin A in the diagnosis of pheochromocytoma and neuroblastoma. *Clin Chim Acta* 1995;239:57–63.
- [40] Stridsberg M, Husebye ES. Chromogranin A and chromogranin B are sensitive circulating markers for pheochromocytoma. *Eur J Endocrinol* 1997;136:67–73.
- [41] Giovannella L, Ceriani L. Serum chromogranin-alpha immunoradiometric assay in the diagnosis of pheochromocytoma. *Int J Biol Markers* 2002;17:130–4.
- [42] d'Herbomez M, Gouze V, Huglo D, Nocaudie M, Pattou F, Proye C, Wemeau JL, Marchandise X. Chromogranin A assay and (131)I-MIBG scintigraphy for diagnosis and follow-up of pheochromocytoma. *J Nucl Med* 2001;42:993–7.
- [43] d'Herbomez M, Forzy G, Bauters C, Tierny C, Pigny P, Carnaille B, Pattou F, Wemeau JL, Rouaix N. An analysis of the biochemical diagnosis of 66 pheochromocytomas. *Eur J Endocrinol* 2007;156:569–75.
- [44] Giovannella L, Squin N, Ghelfo A, Ceriani L. Chromogranin A immunoradiometric assay in diagnosis of pheochromocytoma: comparison with plasma metanephrines and 123I-MIBG scan. *Q J Nucl Med Mol Imaging* 2006;50:344–7.
- [45] Grossrubatscher E, Dalino P, Vignati F, Gambacorta M, Pugliese R, Boniardi M, Rossetti O, Marocchi A, Bertuzzi M, Loli P. The role of chromogranin A in the management of patients with pheochromocytoma. *Clin Endocrinol (Oxf)* 2006;65:287–93.
- [46] Bravo EL. Evolving concepts in the pathophysiology, diagnosis, and treatment of pheochromocytoma. *Endocr Rev* 1994;15:356–68.
- [47] Bravo EL, Tagle R. Pheochromocytoma: state-of-the-art and future prospects. *Endocr Rev* 2003;24:539–53.
- [48] Algeciras-Schimmich A, Preissner CM, Young Jr WF, Singh RJ, Grebe SK. Plasma chromogranin A or urine fractionated metanephrines follow-up testing improves the diagnostic accuracy of plasma fractionated metanephrines for pheochromocytoma. *J Clin Endocrinol Metab* 2008;93:91–5.
- [49] Canale MP, Bravo EL. Diagnostic specificity of serum chromogranin-A for pheochromocytoma in patients with renal dysfunction. *J Clin Endocrinol Metab* 1994;78:1139–44.
- [50] Stridsberg M, Oberg K, Li Q, Engstrom U, Lundqvist G. Measurements of chromogranin A, chromogranin B (secretogranin I), chromogranin C (secretogranin II) and pancreastatin in plasma and urine from patients with carcinoid tumours and endocrine pancreatic tumours. *J Endocrinol* 1995;144:49–59.
- [51] Sanduleanu S, De Bruine A, Stridsberg M, Jonkers D, Biemond I, Haemeeteman W, Lundqvist G, Stockbrugger RW. Serum chromogranin A as a screening test for gastric enterochromaffin-like cell hyperplasia during acid-suppressive therapy. *Eur J Clin Invest* 2001;31:802–11.
- [52] Igaz P, Mullner K, Hargitai B, Igaz I, Tombol Z, Racz K, Tulassay Z. Marked chromogranin A elevation in a patient with bilateral adrenal incidentalomas, and its rapid normalization after discontinuation of proton pump inhibitor therapy. *Clin Endocrinol (Oxf)* 2007;67:805–6.
- [53] Borch K, Stridsberg M, Burman P, Rehfeld JF. Basal chromogranin A and gastrin concentrations in circulation correlate to endocrine cell proliferation in type-A gastritis. *Scand J Gastroenterol* 1997;32:198–202.
- [54] Spadaro A, Ajello A, Morace C, Zirilli A, D'Arrigo G, Luigiano C, Martino F, Bene A, Migliorato D, Turiano S, Ferrau O, Freni MA. Serum chromogranin-A in hepatocellular carcinoma: diagnostic utility and limits. *World J Gastroenterol* 2005;11:1987–90.
- [55] Stridsberg M, Eriksson B, Oberg K, Janson ET. A comparison between three commercial kits for chromogranin A measurements. *J Endocrinol* 2003;177:337–41.
- [56] Bilek R, Safarik L, Ciprova V, Vlcek P, Lisa L. Chromogranin A, a member of neuroendocrine secretory proteins as a selective marker for laboratory diagnosis of pheochromocytoma. *Physiol Res* 2008;57:S171–9.
- [57] Stridsberg M, Eriksson B, Fellstrom B, Kristiansson G, Tiensuu Janson E. Measurements of chromogranin B can serve as a complement to chromogranin A. *Regul Pept* 2007;139:80–3.
- [58] Costeta D, Caliumi C, Alo P, Petramala L, Reale MG, Masciangelo R, Signore A, Cianci R, D'Erasmo E, Letizia C. High plasma levels of human chromogranin A and adrenomedullin in patients with pheochromocytoma. *Tumori* 2005;91:53–8.
- [59] Rao F, Keiser HR, O'Connor DT. Malignant pheochromocytoma. Chromaffin granule transmitters and response to treatment. *Hypertension* 2000;36:1045–52.
- [60] Thompson LD. Pheochromocytoma of the Adrenal gland Scaled Score (PASS) to separate benign from malignant neoplasms: a clinicopathologic and immunophenotypic study of 100 cases. *Am J Surg Pathol* 2002;26:551–66.
- [61] Portela-Gomes GM, Grimelius L, Johansson H, Wilander E, Stridsberg M. Chromogranin A in human neuroendocrine tumors: an immunohistochemical study with region-specific antibodies. *Am J Surg Pathol* 2001;25:1261–7.
- [62] Curry WJ, Shaw C, Johnston CF, Thim L, Buchanan KD. Isolation and primary structure of a novel chromogranin A-derived peptide, WE-14, from a human midgut carcinoid tumour. *FEBS Lett* 1992;301:319–21.
- [63] Conlon JM, Hamberger B, Grimelius L. Isolation of peptides arising from the specific posttranslational processing of chromogranin A and chromogranin B from human pheochromocytoma tissue. *Peptides* 1992;13:639–44.
- [64] Gleeson CM, Curry WJ, Johnston CF, Buchanan KD. Occurrence of WE-14 and chromogranin A-derived peptides in tissues of the human and bovine gastroentero-pancreatic system and in human neuroendocrine neoplasia. *J Endocrinol* 1996;151:409–20.
- [65] Montero-Hadjadje M, Vaudry H, Turquier V, Leprince J, Do Rego JL, Gallo-Payet N, Hadjadj J, Anouar Y. Localization and characterization of evolutionarily conserved chromogranin A-derived peptides in the rat and human pituitary and adrenal glands. *Cell Tissue Res* 2002;310:223–36.
- [66] Anouar Y, Yon L, Guillemot J, Thouénon E, Barbier L, Gimenez-Roqueplo AP, Bertherat J, Lefebvre H, Klein M, Muresan M, Grouzmann E, Plouin PF, Vaudry H, Elkahoul AG. Development of novel tools for the diagnosis and prognosis of pheochromocytoma using peptide marker immunoassay and gene expression profiling approaches. *Ann NY Acad Sci* 2006;1073:533–40.
- [67] Guillemot J, Compagnon P, Cartier D, Thouénon E, Bastard C, Lihmann I, Pichon P, Thuillez C, Plouin PF, Bertherat J, Anouar Y, Kuhn JM, Yon L, Lefebvre H. Metoclopramide stimulates catecholamine- and granin-derived peptide secretion from pheochromocytoma cells through activation of serotonin type 4 (5-HT4) receptors. *Endocr Relat Cancer* 2009;16:281–90.
- [68] Bishop AE, Bretherton-Watt D, Hamid QA, Fahey M, Shepherd N, Valentino K, Tatamoto K, Ghatei MA, Bloom SR, Polak JM. The occurrence of pancreastatin in tumours of the diffuse neuroendocrine system. *Mol Cell Probes* 1988;2:225–35.
- [69] Benedum UM, Lamouroux A, Konecki DS, Rosa P, Hille A, Baeuerle PA, Frank R, Lottspeich F, Mallet J, Huttner WB. The primary structure of human secretogranin I (chromogranin B): comparison with chromogranin A reveals homologous terminal domains and a large intervening variable region. *EMBO J* 1987;6:1203–11.
- [70] Fischer-Colbrie R, Lassmann H, Hagn C, Winkler H. Immunological studies on the distribution of chromogranin A and B in endocrine and nervous tissues. *Neuroscience* 1985;16:547–55.
- [71] Hagn C, Schmid KW, Fischer-Colbrie R, Winkler H. Chromogranin A, B, and C in human adrenal medulla and endocrine tissues. *Lab Invest* 1986;55:405–11.
- [72] Lloyd RV, Cano M, Rosa P, Hille A, Huttner WB. Distribution of chromogranin A and secretogranin I (chromogranin B) in neuroendocrine cells and tumors. *Am J Pathol* 1988;130:296–304.
- [73] Weiler R, Fischer-Colbrie R, Schmid KW, Feichtinger H, Bussolati G, Grimelius L, Krisch K, Kerl H, O'Connor D, Winkler H. Immunological studies on the occurrence and properties of chromogranin A and B and secretogranin II in endocrine tumors. *Am J Surg Pathol* 1988;12:877–84.
- [74] Portela-Gomes GM, Stridsberg M, Grimelius L, Falkmer UG, Falkmer S. Expression of chromogranins A, B, and C (secretogranin II) in human adrenal medulla and in benign and malignant pheochromocytomas. An immunohistochemical study with region-specific antibodies. *APMIS* 2004;112:663–73.
- [75] Benjannet S, Leduc R, Lazure C, Seidah NG, Marcinkiewicz M, Chretien M, GAWK, a novel human pituitary polypeptide: isolation, immunocytochemical localization and complete amino acid sequence. *Biochem Biophys Res Commun* 1985;126:602–9.
- [76] Benjannet S, Leduc R, Adrouche N, Falgout JP, Marcinkiewicz M, Seidah NG, Mbikay M, Lazure C, Chretien M. Chromogranin B (secretogranin I), a putative precursor of two novel pituitary peptides through processing at paired basic residues. *FEBS Lett* 1987;224:142–8.
- [77] Bishop AE, Sekiya K, Salahuddin MJ, Carlei F, Rindi G, Fahey M, Steel JH, Hedges M, Domoto T, Fischer-Colbrie R, Winkler H, Krausz T, Ghatei MA, Bloom SR, Polak JM.

- The distribution of GAWK-like immunoreactivity in neuroendocrine cells of the human gut, pancreas, adrenal and pituitary glands and its co-localisation with chromogranin B. *Histochemistry* 1989;90:475–83.
- [78] Sekiya K, Ghatei MA, Salahuddin MJ, Bishop AE, Hamid QA, Ibayashi H, Polak JM, Bloom SR. Production of GAWK (chromogranin-B 420-493)-like immunoreactivity by endocrine tumors and its possible diagnostic value. *J Clin Invest* 1989;83:1834–42.
- [79] Schober M, Fischer-Colbrie R, Schmid KW, Bussolati G, O'Connor DT, Winkler H. Comparison of chromogranins A, B, and secretogranin II in human adrenal medulla and pheochromocytoma. *Lab Invest* 1987;57:385–91.
- [80] Guillemot J, Barbier L, Thouñnon E, Vallet-Erdtmann V, Montero-Hadjadje M, Lefebvre H, Klein M, Muresan M, Plouin PF, Seidah N, Vaudry H, Anouar Y, Yon L. Expression and processing of the neuroendocrine protein secretogranin II in benign and malignant pheochromocytomas. *Ann NY Acad Sci* 2006;1073:527–32.
- [81] Stridsberg M, Eriksson B, Janson ET. Measurements of secretogranins II, III, V and proconvertases 1/3 and 2 in plasma from patients with neuroendocrine tumours. *Regul Pept* 2008;148:95–8.
- [82] Wiedermann CJ. Secretoneurin: a functional neuropeptide in health and disease. *Peptides* 2000;21:1289–98.
- [83] Fischer-Colbrie R, Kirchmair R, Kähler CM, Wiedermann CJ, Saria A. Secretoneurin: a new player in angiogenesis and chemotaxis linking nerves, blood vessels and the immune system. *Curr Protein Pept Sci* 2005;6:373–85.
- [84] Kirchmair R, Hogue-Angeletti R, Gutierrez J, Fischer-Colbrie R, Winkler H. Secretoneurin—a neuropeptide generated in brain, adrenal medulla and other endocrine tissues by proteolytic processing of secretogranin II (chromogranin C). *Neuroscience* 1993;53:359–65.
- [85] Schmid KW, Kunk B, Kirchmair R, Totsch M, Bocker W, Fischer-Colbrie R. Immunohistochemical detection of secretoneurin, a novel neuropeptide endo-proteolytically processed from secretogranin II, in normal human endocrine and neuronal tissues. *Histochem J* 1995;27:473–81.
- [86] Ischia R, Gasser RW, Fischer-Colbrie R, Eder U, Pagani A, Cubeddu LX, Lovisetti-Scamihorn P, Finkenstedt G, Laslop A, Winkler H. Levels and molecular properties of secretoneurin-immunoreactivity in the serum and urine of control and neuroendocrine tumor patients. *J Clin Endocrinol Metab* 2000;85:355–60.
- [87] Anouar Y, Jegou S, Alexandre D, Lihmann I, Conlon JM, Vaudry H. Molecular cloning of frog secretogranin II reveals the occurrence of several highly conserved potential regulatory peptides. *FEBS Lett* 1996;394:295–9.
- [88] Anouar Y, Desmoucelles C, Yon L, Leprince J, Breault L, Gallo-Payet N, Vaudry H. Identification of a novel secretogranin II-derived peptide (SgII(187–252)) in adult and fetal human adrenal glands using antibodies raised against the human recombinant peptide. *J Clin Endocrinol Metab* 1998;83:2944–51.
- [89] Montero-Hadjadje M, Pelletier G, Yon L, Li S, Guillemot J, Magoul R, Tillet Y, Vaudry H, Anouar Y. Biochemical characterization and immunocytochemical localization of EM66, a novel peptide derived from secretogranin II, in the rat pituitary and adrenal glands. *J Histochem Cytochem* 2003;51:1083–95.
- [90] Guillemot J, Ait-Ali D, Turquier V, Montero-Hadjadje M, Fournier A, Vaudry H, Anouar Y, Yon L. Involvement of multiple signaling pathways in PACAP-induced EM66 secretion from chromaffin cells. *Regul Pept* 2006;137:79–88.
- [91] Yon L, Guillemot J, Montero-Hadjadje M, Grumolato L, Leprince J, Lefebvre H, Contesse V, Plouin PF, Vaudry H, Anouar Y. Identification of the secretogranin II-derived peptide EM66 in pheochromocytomas as a potential marker for discriminating benign versus malignant tumors. *J Clin Endocrinol Metab* 2003;88:2579–85.
- [92] Guillemot J, Anouar Y, Montero-Hadjadje M, Grouzmann E, Grumolato L, Roshmaninho-Salgado J, Turquier V, Duparc C, Lefebvre H, Plouin PF, Klein M, Muresan M, Chow BK, Vaudry H, Yon L. Circulating EM66 is a highly sensitive marker for the diagnosis and follow-up of pheochromocytoma. *Int J Cancer* 2006;118:2003–12.
- [93] Buffa R, Pelagi M, Siccardi AG, Curzio M, Yu JY, Bordi C. Identification of the endocrine cells detected by the monoclonal antibody H1SL-19 in human tissues. *Basic Appl Histochem* 1990;34:259–68.
- [94] Shimizu K, Namimatsu S, Kitagawa W, Akasu H, Takatsu K, Sugisaki Y, Tanaka S. Immunohistochemical, biochemical and immunoelectron microscopic analysis of antigenic proteins on neuroendocrine cell tumors using monoclonal antibody H1SL-19. *J Nippon Med Sch* 2002;69:365–72.
- [95] Iguchi H, Natori S, Nawata H, Kato K, Ibayashi H, Chan JS, Seidah NG, Chretien M. Presence of the novel pituitary protein “7B2” in bovine chromaffin granules: possible co-release of 7B2 and catecholamine as induced by nicotine. *J Neurochem* 1987;49:1810–4.
- [96] Brayton KA, Aimi J, Qiu H, Yazdanparast R, Ghatei MA, Polak JM, Bloom SR, Dixon JE. Cloning, characterization, and sequence of a porcine cDNA encoding a secreted neuronal and endocrine protein. *DNA* 1988;7:713–9.
- [97] Marcinkiewicz M, Benjannet S, Falgout JP, Seidah NG, Schurch W, Verdy M, Cantin M, Chretien M. Identification and localization of 7B2 protein in human, porcine, and rat thyroid gland and in human medullary carcinoma. *Endocrinology* 1988;123:866–73.
- [98] Hacker GW, Bishop AE, Terenghi G, Varnedell IM, Aghahowa J, Pollard K, Thurner J, Polak JM. Multiple peptide production and presence of general neuroendocrine markers detected in 12 cases of human pheochromocytoma and in mammalian adrenal glands. *Virchows Arch Pathol Anat Histopathol* 1988;412:399–411.
- [99] Vieau D, Linaud CG, Mbikay M, Lenne F, Chretien M, Luton JP, Bertagna X. Expression of the neuroendocrine cell marker 7B2 in human ACTH secreting tumours. *Clin Endocrinol* 1992;36:597–603.
- [100] Natori S, Iguchi H, Ohashi M, Nawata H. Plasma 7B2 (a novel pituitary protein) immunoreactivity concentrations in patients with various endocrine disorders. *Endocrinol Jpn* 1988;35:651–4.
- [101] Fischer-Colbrie R, Eder S, Lovisetti-Scamihorn P, Becker A, Laslop A. Neuroendocrine secretory protein 55: a novel marker for the constitutive secretory pathway. *Ann NY Acad Sci* 2002;971:317–22.
- [102] Bauer R, Weiss C, Marksteiner J, Doblinger A, Fischer-Colbrie R, Laslop A. The new chromogranin-like protein NESP55 is preferentially localized in adrenaline-synthesizing cells of the bovine and rat adrenal medulla. *Neurosci Lett* 1999;263:13–6.
- [103] Jakobsen AM, Ahlman H, Kolby L, Abrahamsson J, Fischer-Colbrie R, Nilsson O. NESP55, a novel chromogranin-like peptide, is expressed in endocrine tumours of the pancreas and adrenal medulla but not in ileal carcinoids. *Br J Cancer* 2003;88:1746–54.
- [104] Nilsson O, Jakobsen AM, Kolby L, Bernhardt P, Forssell-Aronsson E, Ahlman H. Importance of vesicle proteins in the diagnosis and treatment of neuroendocrine tumours. *Ann NY Acad Sci* 2004;1014:280–3.
- [105] Srivastava A, Padilla O, Fischer-Colbrie R, Tischler AS, Dayal Y. Neuroendocrine secretory protein-55 (NESP-55) expression discriminates pancreatic endocrine tumors and pheochromocytomas from gastrointestinal and pulmonary carcinoids. *Am J Surg Pathol* 2004;28:1371–8.
- [106] Mzhavia N, Qian Y, Feng Y, Che FY, Devi LA, Fricker LD. Processing of proSAAS in neuroendocrine cell lines. *Biochem J* 2002;361:67–76.
- [107] Chakraborty TR, Tkalych O, Nanno D, Garcia AL, Devi LA, Salton SR. Quantification of VGF- and pro-SAAS-derived peptides in endocrine tissues and the brain, and their regulation by diet and cold stress. *Brain Res* 2006;1089:21–32.
- [108] Rindi G, Licini L, Necchi V, Bottarelli L, Campanini N, Azzoni C, Favret M, Giordano G, D'Amato F, Brancia C, Solcia E, Ferri GL. Peptide products of the neurotrophin-inducible gene vgf are produced in human neuroendocrine cells from early development and increase in hyperplasia and neoplasia. *J Clin Endocrinol Metab* 2007;92:2811–5.
- [109] Feng Y, Reznik SE, Fricker LD. Distribution of proSAAS-derived peptides in rat neuroendocrine tissues. *Neuroscience* 2001;105:469–78.
- [110] Tischler AS. Molecular and cellular biology of pheochromocytomas and extra-adrenal paragangliomas. *Endocr Pathol* 2006;17:321–8.
- [111] Pacak K, Eisenhofer G. An assessment of biochemical tests for the diagnosis of pheochromocytoma. *Nat Clin Pract Endocrinol Metab* 2007;3:744–5.
- [112] Gao B, Meng F, Bian W, Chen J, Zhao H, Ma G, Shi B, Zhang J, Liu Y, Xu Z. Development and validation of pheochromocytoma of the adrenal gland scaled score for predicting malignant pheochromocytomas. *Urology* 2006;68:282–6.
- [113] Strong VE, Kennedy T, Al-Ahmadie H, Tang L, Coleman J, Fong Y, Brennan M, Ghossein RA. Prognostic indicators of malignancy in adrenal pheochromocytomas: clinical, histopathologic, and cell cycle/apoptosis gene expression analysis. *Surgery* 2008;143:759–68.
- [114] Brouwers FM, Elkahloun AG, Munson PJ, Eisenhofer G, Barb J, Linehan WM, Lenders JW, De Krijger R, Mannelli M, Udelsman R, Ocal IT, Shulkin BL, Bornstein SR, Breza J, Ksinantova L, Pacak K. Gene expression profiling of benign and malignant pheochromocytoma. *Ann NY Acad Sci* 2006;1073:541–56.
- [115] Eisenhofer G, Goldstein DS, Sullivan P, Csako G, Brouwers FM, Lai EW, Adams KT, Pacak K. Biochemical and clinical manifestations of dopamine-producing paragangliomas: utility of plasma methoxytyramine. *J Clin Endocrinol Metab* 2005;90:2068–75.
- [116] Thouñnon E, Pierre A, Guillemot J, Yon L, Eisenhofer G, Anouar Y. Genetic markers for the diagnosis and prognosis of pheochromocytoma. *Expert Rev Endocrinol Metab* 2009;4:45–52.
- [117] Neumann HP. Imaging vs biochemical testing for pheochromocytoma. *Jama* 2002;288:314–5.
- [118] Sawka AM, Gafni A, Thabane L, Young Jr WF. The economic implications of three biochemical screening algorithms for pheochromocytoma. *J Clin Endocrinol Metab* 2004;89:2859–66.
- [119] Tischler AS. Pheochromocytoma and extra-adrenal paraganglioma. *Updates Arch Pathol Lab Med* 2008;132:1272–84.
- [120] Kimura N, Watanabe T, Noshiro T, Shizawa S, Miura Y. Histological grading of adrenal and extra-adrenal pheochromocytomas and relationship to prognosis: a clinicopathological analysis of 116 adrenal pheochromocytomas and 30 extra-adrenal sympathetic paragangliomas including 38 malignant tumors. *Endocr Pathol* 2005;16:23–32.
- [121] Bilek R, Safarik L, Ciprová V, Vlcek P, Lisá L. Chromogranin A, a member of neuroendocrine secretory proteins as a selective marker for laboratory diagnosis of pheochromocytoma. *Physiol Res* 2008;57:S171–9.
- [122] Colombo B, Curnis F, Foglieni C, Monno A, Arrigoni G, Corti A. Chromogranin A expression in neoplastic cells affects tumor growth and morphogenesis in mouse models. *Cancer Res* 2002;62:941–6.